

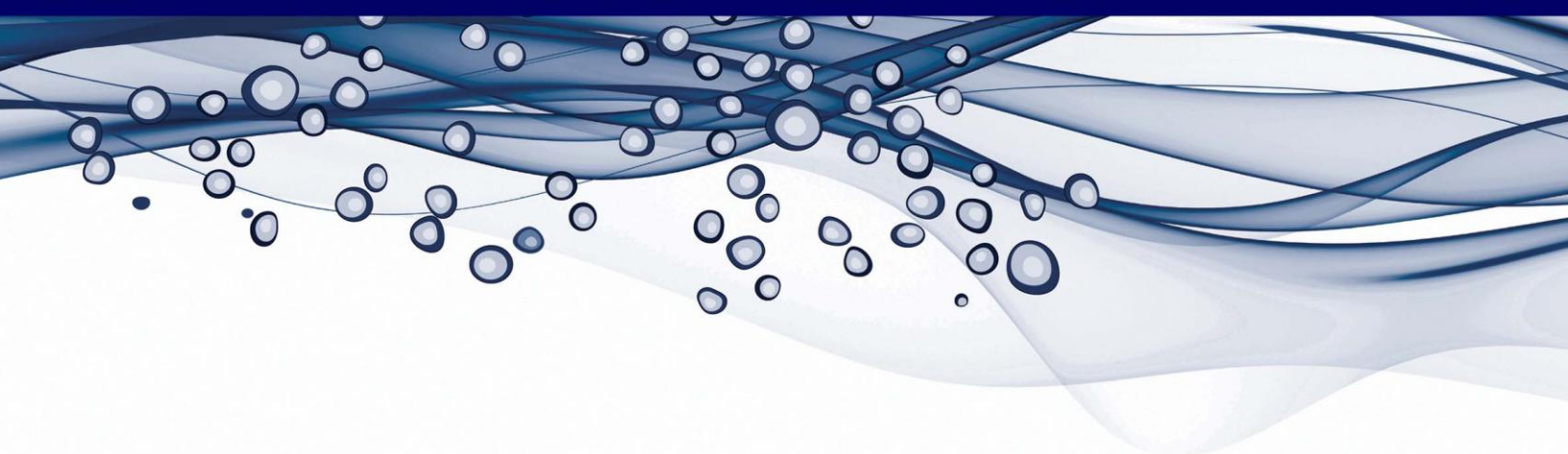
# Cellometer<sup>®</sup>

**Product Number:** CS0-0110-500ML

**Description:** Cellometer ViaStain<sup>™</sup> Yeast Dilution Buffer

**Instrument (s):** Cellometer X2, Vision10x

## Instruction Booklet: ViaStain<sup>™</sup> Yeast Dilution Buffer



This product is for RESEARCH USE ONLY and is not approved for diagnostic or therapeutic use.

8001308 Rev. A



## Product

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Part Number: **CS0-0110-500ML**

Description: Cellometer ViaStain™ Yeast Dilution Buffer

Manufacturer Lot Number: **YYMMDD-RR-B**

Volume: **500 mL**

pH Range: ≤ 10.0

## Description

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The ViaStain™ Yeast Dilution Buffer in combination with the ViaStain™ Yeast Live / Dead Cell staining solution enables the user to quantitatively distinguish live and dead yeast in pure cultures and in cultures containing debris such as corn mash using the Cellometer system. The Dilution Buffer is used to dilute the yeast sample in order to promote and optimize live / dead fluorescent staining of the yeast cells.

## Materials

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### Materials Supplied

1. Cellometer ViaStain™ Yeast Dilution Buffer (CS0-0110-500ML)\*

\*One 500 mL bottle of Yeast Dilution Buffer

### Materials Required

1. Micro centrifuge tube
2. Pipettor
3. Cellometer counting chamber (SD100 or PD100)
4. Cellometer Vision10x (with Fluorescence Optical Module F101, VB-535-401, or equivalent and F304, VB-660-501, or equivalent) or a Cellometer X2

## Procedure

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1. Dilute corn mash sample 10-fold (by weight) in water and mix well.
2. Allow corn mash debris in diluted sample to settle for 30 sec.
3. Pipette 10 µL of diluted corn mash (while avoiding large debris in the mixture) and dispense into a microcentrifuge tube.
4. Add 10 µL of yeast dilution buffer to microcentrifuge tube and mix well by pipetting up and down 3 times.
5. Add 20 µL ViaStain™ Yeast Live / Dead Cell staining solution to microcentrifuge tube and mix well by pipetting up and down 3 times.
6. Incubate the stained corn sample for 1-2 min at room temperature.
7. Pipette sample up and down 3 times to mix and then load 20 µL into a counting chamber (if using SD100 slides, peel plastic film off both sides before loading).

8. Place loaded slide on a Kimwipe® and wait 1 min before inserting sample into instrument to allow sample to settle in the chamber.
9. Select the appropriate assay type for yeast viability measurement.
10. Preview brightfield and fluorescent images.
11. Count.

### Storage and Handling

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Store the Yeast Dilution Buffer at room temperature. The ViaStain™ Yeast Dilution Buffer is a buffer at high pH; safety precautions must be taken when handling the solution. Please consult the Material Safety Data Sheet for more safety information, found on [www.nexcelom.com/Products](http://www.nexcelom.com/Products).

### Warranty

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This product is for RESEARCH USE ONLY and is not approved for diagnostic or therapeutic use. Product is warranted to meet the specifications outlined in the Certificate of Analysis when stored and used according to the manufacturer's instructions. No other warranty, expressed or implied (such as merchantability, fitness for a particular purpose, or non-infringement) is granted. Warranty is valid until the expiration date stated on the product label. If no expiration is listed, the warranty is valid for 12 months from the date of product receipt.

Warranty will be void if product is stored incorrectly, the recommended protocol is not followed, or the product is used for a different application.

### Quality Control

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Manufactured and tested according to SOP #: 8001434

Passed



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Matthew Bularzik, Quality Engineer

03/10/2014